

Development of PSMA x CD3 T-cell engagers using an integrated, functional approach

BACKGROUND

PSMA-targeted bispecifics for prostate cancer

CD3 T-cell engagers (TCEs) targeting prostate-specific membrane antigen (PSMA) have emerged as a promising approach for the treatment of metastatic castrationresistant prostate cancer.¹ However, doselimiting toxicities have been a barrier to bringing them to patients.²

Identifying TCEs that balance anti-tumor potency with potential toxicities requires simultaneous tuning of both the CD3- and tumor-binding arm. To develop optimal TCE candidates, we engineer hundreds of bispecific molecules from highly diverse parental antibodies, and employ a high-throughput process to identify molecules with desired functional properties.³

Fine-tune T-cell engagers to decouple tumor-cell killing and cytokine release

Here, we show a function-first approach to address the clinical challenges observed in PSMA-targeted bispecifics. Starting with a diverse panel of more than 180 PSMA x CD3 bispecifics,³ our aim was to identify TCEs against PSMA that:

- show potent tumor-cell killing, with low levels of cytokine release
- induce T-cell properties associated with anti-tumor immune responses, including proliferation of CD4⁺ and CD8⁺ T-cell subsets
- have favorable developability profiles

OUTCOME

PSMA x CD3 T-cell engagers that induce potent tumor- cell killing with reduced cytokine release

Starting with highly diverse CD3- and PSMAbinding antibodies, we used a functional approach to identify promising TCE candidates. These TCEs show significantly lower cytokine release than clinical benchmarks, while maintaining potent killing of PSMA-expressing cancer cell lines in vitro.

In parallel, we are investigating novel CD3-binding arms that can further decouple tumor-cell killing and cytokine release.

Additional preclinical characterization will determine the potential of these PSMAtargeted bispecifics as therapeutics for prostate cancer.

PSMA x CD3 bispecifics that are differentiated from clinical benchmarks

T-cell engagers with optimal pairing of CD3- and PSMA-binders induce potent tumor-cell killing and low cytokine release





Robust T cell-mediated immune responses



Figure 2. T-cell profiling in cancer cell lines with high and low PSMA expression. bsAb 1 and 2 induce proliferation of CD4⁺ and CD8⁺ T-cell subsets, production of T-cell redirection chemokine (CXCL10) and activation cytokines (IL-12), and release of granzyme B from lymphocytes, at levels comparable to AMG 160. PBMC effector and target cells were incubated at a ratio of 10:1 for 48 hours, with markers measured using Meso Scale Discovery[®] and flow cytometry. Results from a single representative PBMC donor are shown.

REFERENCES

- 1. Simão DC, et al. (2023). Cancers (Basel). 15(5):1412. doi: 10.3390/cancers15051412. 2. Heitmann JS, et al. (2021). Cancers (Basel). 13(3):549. doi: 10.3390/cancers13030549.
- 3. Mai J, et al. (2023). J Immunother Cancer. 11(Issue Suppl 1):1367. doi: 10.1136/jitc-2023-SITC2023.1367.
- 4. Deegen P, et al. (2021). Clin Cancer Res. 27(10):2928-2937. doi: 10.1158/1078-0432.CCR-20-3725.

5. Dang K, et al. (2021). J Immunother Cancer. 9:e002488. doi: 10.1136/jitc-2021-002488. 6. Zekri L. et al. (2021). EMBO Mol Med. 13(2):e11902. doi: 10.15252/emmm.201911902 7. Gottschalk R, et al. (2021). Cancer Res. 81 (13_Suppl): LB172. doi: 10.1158/1538-7445.AM2021-LB172. 8. Nisthal A, et al. (2020). Cancer Res. 80 (16_Suppl): 5663. doi: 10.1158/1538-7445.AM2020-5663.

AUTHORS Valentine de Puyraimond, Matt Mai, Alaa Amash, Nathalie Blamey, Gabrielle Conaghan, Jessica Fernandes Scortecci, Allie Goodman, Ahn Lee, Irene Yu, Franziska von Bank, Kate Caldwell, Lauren Clifford, Peter Bergqvist, Kelly Bullock, Melissa Cid, Cindy-Lee Crichlow, Lindsay Devorkin, Fiona Dickson, Patrick Farber, Stefan Hannie, Ingrid Knarston, Courteney Lai, Vivian Li, Stephanie Masterman, Iwona Niemietz, Philippe Pouliot, Ping Xiang, Christopher Williamson, Bryan C. Barnhart, Raffi Tonikian* *presenter AUTHOR AFFILIATION AbCellera, Vancouver, Canada

6	CD3 arm	TAA arm
	CD3-1	PSMA-1
	CD3-2	PSMA-1

Figure 1. Selection of PSMA x CD3 bispecifics with functional activity between the range of two clinical benchmarks. (A) Two PSMA x CD3 bispecifics (bsAb 1 and 2) show max tumor-cell killing that is comparable to clinical benchmark AMG 160,⁴ with reduced cytokine release. Function was assessed with a high-throughput T-cell-dependent cellular cytotoxicity (TDCC) assay using human peripheral blood mononuclear cells (PBMCs) incubated with target cells at a 10:1 ratio for 72 hours. (B) The TAA arm shared by bsAb 1 and 2 shows membrane-proximal binding to PSMA. The structure was generated using a size-exclusion chromatography-purified complex and cryo-electron microscopy. (C) CD3-binding arms were selected from AbCellera's diverse CD3 panel, which is visualized using Celium™ and colored by binding competition with the commonly used CD3-binding antibody SP34-2. The CD3 arms used for bsAb 1 and 2 bind epitopes that are

T-cell-mediated lysis of PSMA-expressing prostate cancer cells

Potent tumor-cell killing and reduced cytokine release is consistent across cell lines and donors





Next steps



antibodies that generate TCEs with high potency and reduced cytokine release across multiple tumor targets. These were combined with three PSMA-binding antibodies and assessed by high-throughput TDCC. Our panel with unique CD3-binders shows enrichment for TCEs with desired functional profiles; several candidates will be assessed in parallel with bsAb 1 and 2. (B) Tumor-cell killing and cytokine release for an example bispecific (bsAb 3) from this panel is shown.

6359

DOWNLOAD POSTER

Figure 3. Tumor-cell killing in human and cyno PSMA-expressing cancer cells. (A) Tumor-cell killing activity of bsAb 1 and 2 is consistent across varied levels of PSMA expression in multiple human cancer cell lines. (B) Cytokine release and tumor-cell killing profiles were consistent across cell lines and between PBMC donors, measured using the assay described in Figure 1A with a 72–96 hour incubation. Max cytokines (pg/mL) were standardized per condition (donor and cell line) by subtracting the mean and dividing by the standard deviation. Dose-dependent response curves from a representative donor are shown in Figure 1A. (C) bsAb 1 and 2 show dose-dependent killing of cyno PSMA-expressing cells with lower cytokine release than AMG 160. Cyno pan T-cell donor cells were incubated with target cells at a 10:1 ratio for 48 hours. Data not shown for IL-6, which showed low concentration across bsAb 1/2 and benchmarks.